



EDGEWOOD

CHEMICAL BIOLOGICAL CENTER

U.S. ARMY SOLDIER AND BIOLOGICAL CHEMICAL COMMAND

ECBC-TR-255

**BACKGROUND AEROSOLS AND BW DETECTION:
PROBLEMS AND SUGGESTED SOLUTIONS**

**Amnon Birenzvige
Charles Wick**

RESEARCH AND TECHNOLOGY DIRECTORATE

September 2002

**Approved for public release;
distribution is unlimited.**



Aberdeen Proving Ground, MD 21010-5424

20030110 026

Disclaimer

The findings in this report are not to be construed as an official Department of the Army position unless so designated by other authorizing documents.

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave Blank)	2. REPORT DATE	3. REPORT TYPE AND DATES COVERED
	2002 September	Final; 02 Mar - 02 Jun
4. TITLE AND SUBTITLE		5. FUNDING NUMBERS
Background Aerosols and BW Detection: Problems and Suggested Solutions		PR-206023
6. AUTHOR(S)		
Birenzvige, Ammon; and Wick, Charles		
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)		8. PERFORMING ORGANIZATION REPORT NUMBER
DIR, ECBC, ATTN: AMSSB-RRT-II, APG, MD 21010-5424		ECBC-TR-255
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES)		10. SPONSORING/MONITORING AGENCY REPORT NUMBER
11. SUPPLEMENTARY NOTES		
12a. DISTRIBUTION/AVAILABILITY STATEMENT		12b. DISTRIBUTION CODE
Approved for public release; distribution is unlimited.		
13. ABSTRACT (Maximum 200 words)		
<p>Detection of Biological Warfare (BW) Agents is a formidable task, exacerbated by the fact that some microbes can be highly infective. Thus, only a few microbes delivered to an appropriate site can cause disease. Because of this issue, detectors need to have very high sensitivity. To further complicate things, the atmosphere frequently contains abundant biological and nonbiological particles that can interfere with the detection process, thus creating false alarms. In this report, the authors briefly review the difficulties that background aerosols cause in the detection process and suggest possible ways of alleviating these difficulties.</p>		
14. SUBJECT TERMS		15. NUMBER OF PAGES
Bioaerosol Background aerosol	BW Agents BW Detection	Data fusion
		20
		16. PRICE CODE
17. SECURITY CLASSIFICATION OF REPORT UNCLASSIFIED	18. SECURITY CLASSIFICATION OF THIS PAGE UNCLASSIFIED	19. SECURITY CLASSIFICATION OF ABSTRACT UNCLASSIFIED
		20. LIMITATION OF ABSTRACT UL

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)
Prescribed by ANSI Std. Z39-18
298-102

Blank

PREFACE

The work described in this report was performed under Project No. 206023. This work was started in March 2002 and completed in June 2002.

The use of either trade or manufacturers' names in this report does not constitute an official endorsement of any commercial products. This report may not be cited for purposes of advertisement.

This report has been approved for public release. Registered users should request additional copies from the Defense Technical Information Center; unregistered users should direct such requests to the National Technical Information Center.

Blank

CONTENTS

1.	INTRODUCTION	7
2.	BW DETECTION AND THE BACKGROUND - PROBLEM DEFINITION	7
3.	DISCUSSION	11
4.	CONCLUSIONS AND RECOMMENDATIONS	16
	LITERATURE CITED	19

FIGURES

1.	Pictorial Illustration of the Background and the BW Detection Problem	8
2.	Pictorial Illustration Comparing Minimum Detection Level with Common Background	9
3.	Example of Bioaerosols Found in Nature	10
4.	Example of Particle Counts (Number Concentration) During the Day	12
5.	Example of the Spatial Distribution of Environmental Aerosols.....	12
6.	Example of Cultureable Bacteria Concentration in the Air	13
7.	Example of a Py-GC/IMS (Developmental BW Detector at ECBC) Signature in the Background	14
8.	Example of the SPFA Signature in the Background	15

BACKGROUND AEROSOLS AND BW DETECTION: PROBLEMS AND SUGGESTED SOLUTIONS

1. INTRODUCTION

The background atmosphere contains biological and nonbiological material that can interfere with detecting Biological Warfare (BW) agents. These Bioaerosols are comprised of bacteria, viruses, spores, pollen, fungi, insect parts, etc.^{1,2,3} These rich diversifications of microbes also contain all the microbe breakdown products that are in the process of further decay. In addition, not all the microbes are viable, and many of them available for collection are not named, and thus are unknowns. This situation is made more complicated by their continual variation in concentration in time and space, a phenomena that is not well understood. Some of the factors affecting the background include: time of day, season, local meteorology, human activities, land use, geographic location, and a host of other, as yet, unknown factors.

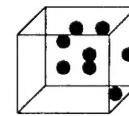
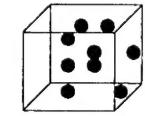
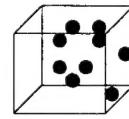
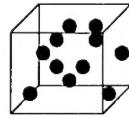
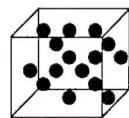
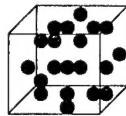
2. BW DETECTION AND THE BACKGROUND - PROBLEM DEFINITION

Figure 1 illustrates the problem the background presents for BW detection. Close to the BW dissemination point, the number concentration of the BW agent particles (red dots in Figure 1) is larger than the number concentration of the background aerosol particles (green dots in Figure 1). Hence, detection is not a problem. As the BW cloud travels down wind and disperses, the background concentration remains the same (more or less), while the number concentration of the agent particles diminishes. The same problem is also illustrated in Figure 2. As noted in Figure 2, usually the desired sensitivity for BW detection falls within the background concentration. To put this another way, when the BW agent can be seen by the eye and the means of dispersal observed, identifying and detecting it as a biological agent is easier to determine. As the material disperses into the atmosphere and floats its way down wind, the concentration diminishes. In many situations, as soon as the material mixes with the indigenous microbes, it becomes less and less concentrated. Recall, for a moment, that only a few microbes may cause infection, and this condition forms the nature of the problem faced by detector systems - low concentration of highly infective microbes that are greatly outnumbered by the native microbes. Separating these different microbes from each other, or more directly separating the target microbes from the background microbes for detection and identification is the real nature of the task presented.

Figure 3 shows examples of biological aerosols that can be found in the background atmosphere. These aerosols can range in size from a fraction of micrometer to several tens of microns. This figure represents several issues regarding the difficulty of separating target microbes from the rest of the mix. First, some of these are found all over the world. Some species of fungi have vegetative parts that when broken apart are similar in size to their spores, both sexual and asexual to make it more interesting. Some of the bacteria have

*Note that BW agents are aerosol particles, in contrast to CW agents that can be either gas (vapor), liquid, or solid aerosols, depending on the agent, on the ambient temperature, and on the dissemination method.

Detection Problem and the Background

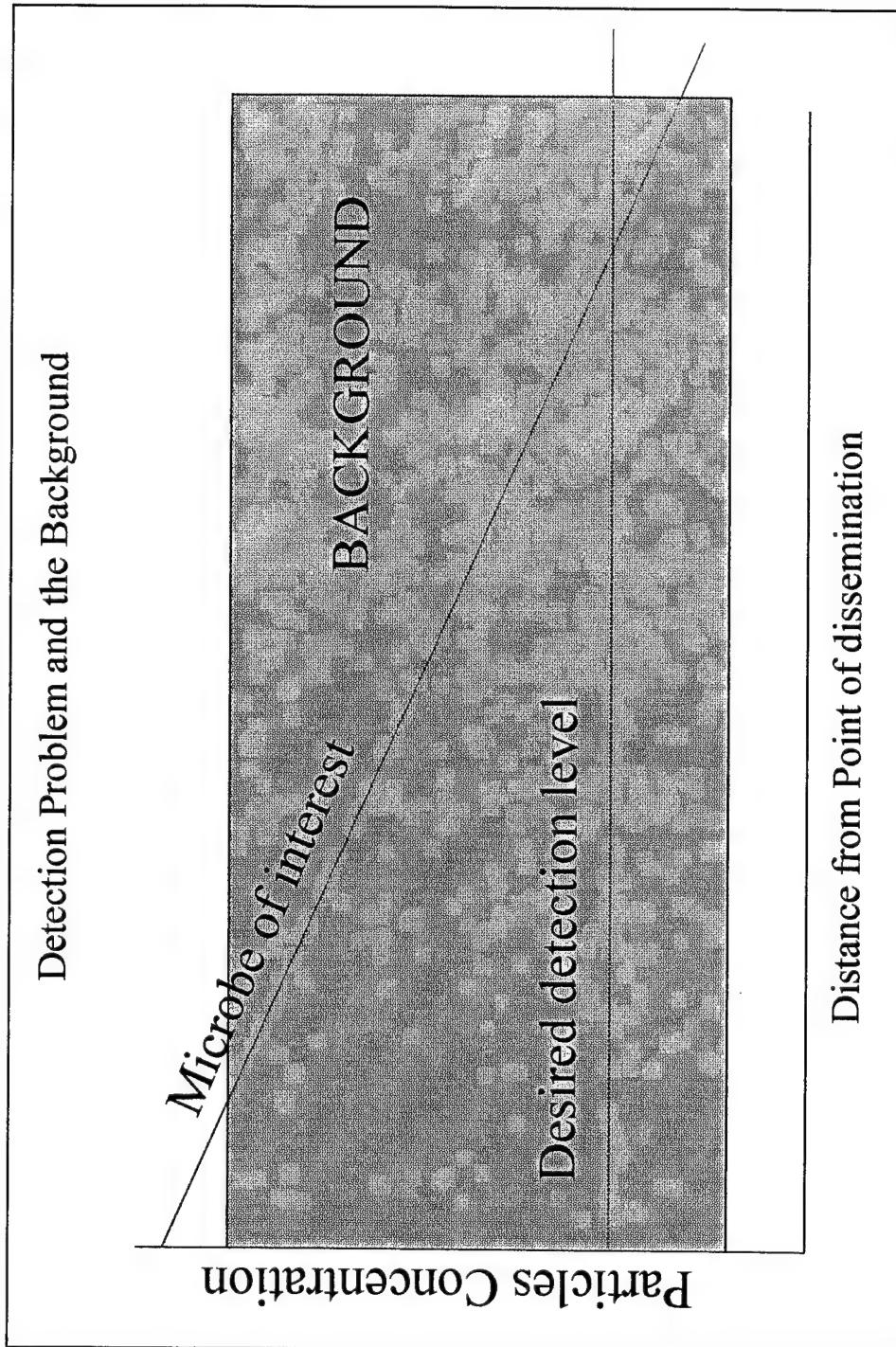


- Microbe of interest
- Background particle

As the distance from the source increases, the background becomes more predominate

Figure 1. Pictorial Illustration of the Background and the BW Detection Problem

Detection Problem and the Background



Distance from Point of dissemination

Figure 2. Pictorial Illustration Comparing Minimum Detection Level with Common Background

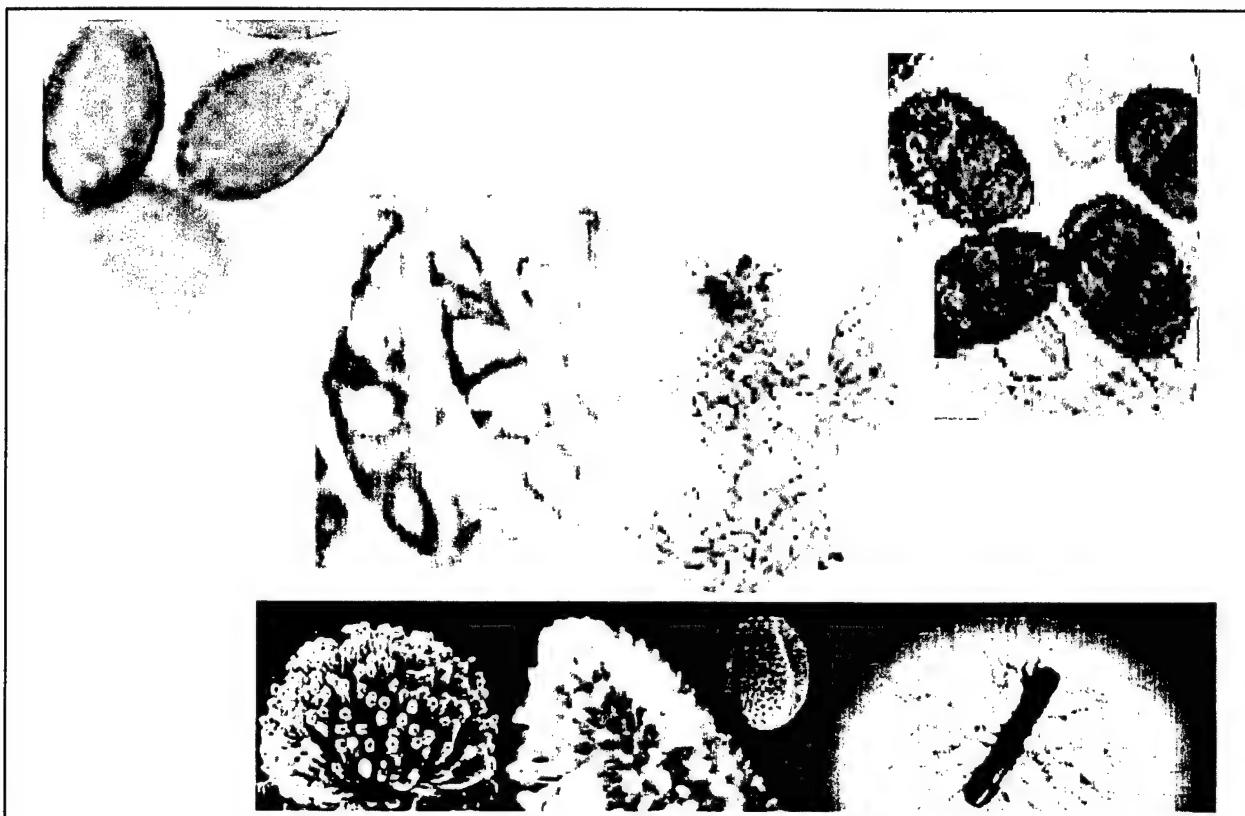


Figure 3. Example of Bioaerosols Found in Nature.
(Picture provided by courtesy of Dr. Charles Wick, ECBC)

projections that can also dry and break apart, forming smaller particles. Combined with enormous numbers of these microbes, the detection and separation challenges begin to take shape.

Various factors affect the background concentration of nonbiological and biological aerosols. Biological aerosols exhibit diurnal and seasonal cycles. The concentration of both depends on geography and land use (desert, urban, rural, forest, agricultural activity, etc.), human activities, etc. Some of the variability is not well understood, and only small fractions of the biological species have been identified. This does not imply that the issue of sampling, detecting, and identifying requires that all competing microbes be identified and cataloged. Not at all. What is implied is a need to develop and test our collectors and detectors under natural real world conditions.

3.

DISCUSSION

Sources of biological aerosol are both natural and man-made. Soil and plants are a major source of bioaerosols.^{4,5,6} Human activities contribute greatly to the atmospheric aerosol and bioaerosol load (e.g., vehicular traffic results in resuspension of particles, and water treatment plants are a good source of bioaerosols of various types).

Figures 4-8 show several examples of background information collected by different means and at different times and locations. Figure 4 shows an example of how the background particle number concentrations can vary within very short periods. As can be seen, the number concentration of the particles can more than double within a few minutes. Figure 5 illustrates how the particle concentration can vary spatially within short distances of several hundreds of meters.*

Figure 6 shows the number concentration of cultureable bacteria particles. Again, the number concentration of particles containing cultureable bacteria can vary by a factor of 2 or more over a relatively short time-span. It must be emphasized that bacterial particles comprise only a small fraction of the total bioaerosols. In addition, only a small fraction of the total bacteria can be cultured.

Figures 7 and 8 show the response of two developmental BW detection systems in the outdoor background. Figure 7 shows the response of a Pyrolysis Gas Chromatograph/Ion Mobility Spectrometer (Py/GC-IMS) that is under development at ECBC.⁷ In this detector, particles are collected and deposited onto a quartz wire over several minutes (nominally 3-5 min). Following the collection, the material collected on the wire is pyrolyzed by passing current through the wire and raising its temperature to 350 °C within 5 sec. As a result of the Pyrolysis, biological material produces various semivolatile compounds. The mixture of vapors is separated to its components by passing through a chromatographic column into a time of flight ion mobility spectrometer after passing through an ionizing volume. Some classes of bioaerosols give characteristic signatures. For example, gram positive spores produce Picolinic acid that is a decomposition product of dipicolinic acid.⁸ Work is now in progress to determine whether other classes of bioaerosols also have typical signatures.

Figure 8 shows an example of data collected outdoors by the Single Particle Fluorescence Analyzer (SPFA), a developmental detector at the Naval Research Laboratory (NRL), Washington, DC.^{9,10,11} The SPFA counts and sizes all entering particles using a cw laser for optical scattering. In addition, it is designed to differentiate particles of biological origin (also known as bioaerosols) from all other aerosols. It achieves this by using a second pulsed laser at a 266 nm wavelength to excite UV fluorescence from each biological particle as they pass by. Because certain amino acids (principally tryptophan and tyrosine) fluoresce when excited near their absorption peaks in the UV region of the spectrum, a strong particle fluorescence signal indicates the presence of biological material, since these amino acids occur in nearly all proteins. Therefore, this spectroscopic characteristic was used to indicate the presence of aerosols of biological origin.^{12,13,14}

*Birenvigie, A., Research and Technology Directorate, U.S. Army Edgewood Chemical Biological Center, Aberdeen Proving Ground, MD, 1999, unpublished data.

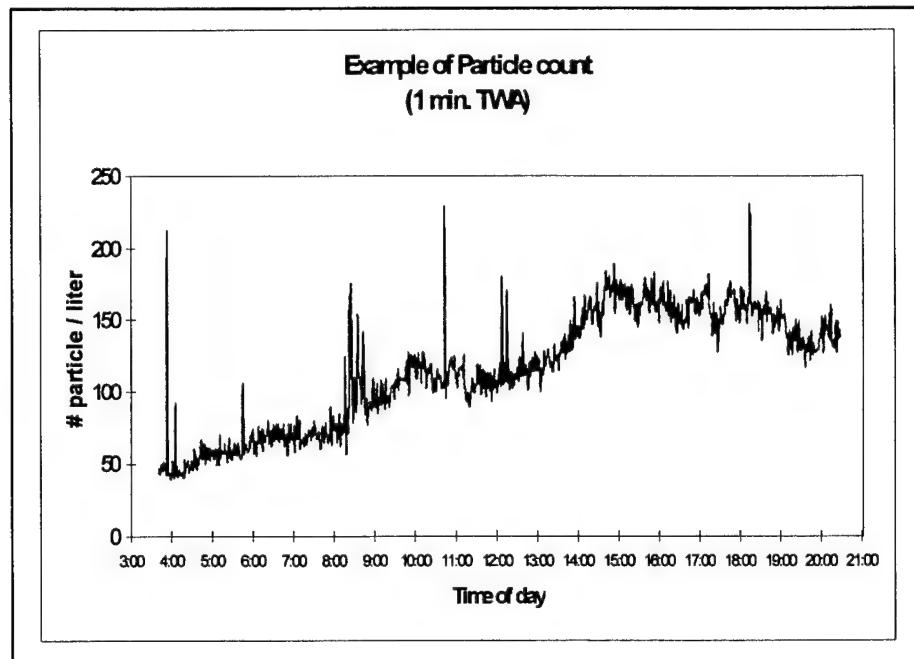


Figure 4. Example of Particle Counts (Number Concentration) During the Day

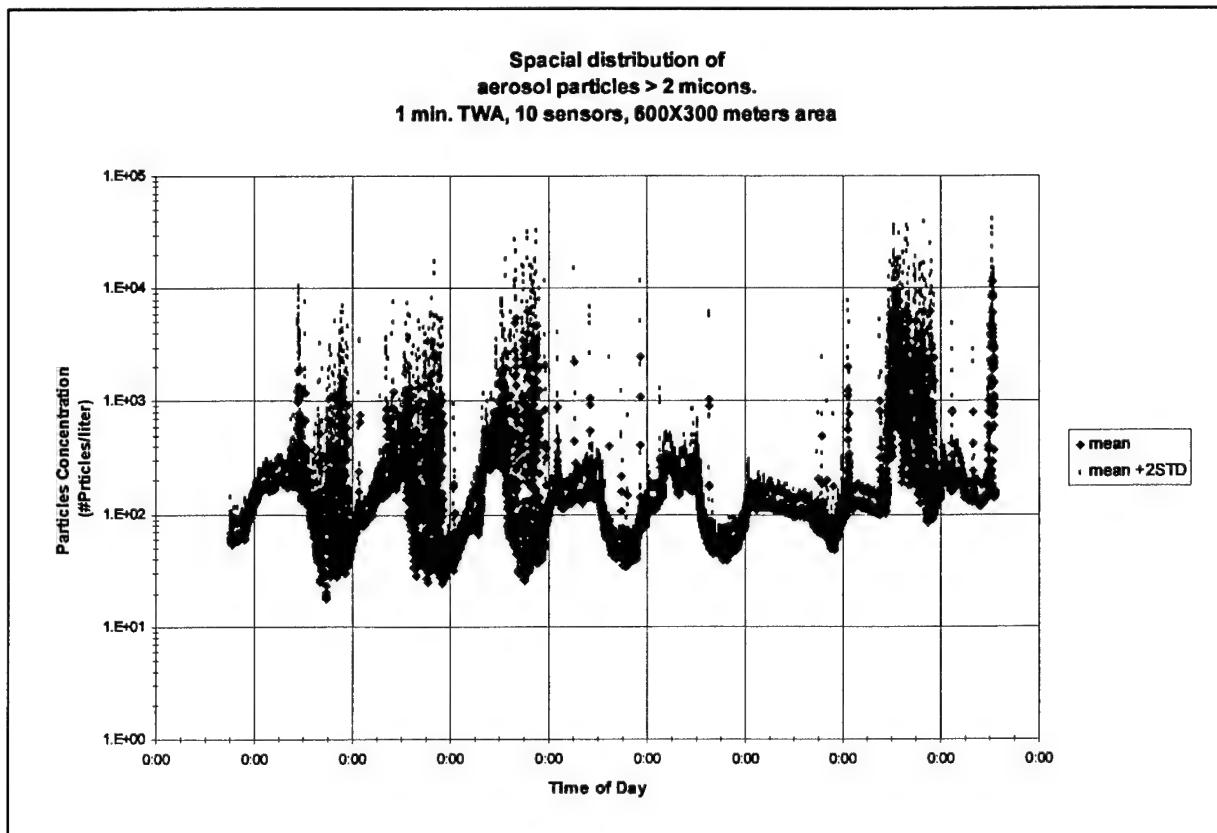


Figure 5. Example of the Spatial Distribution of Environmental Aerosols

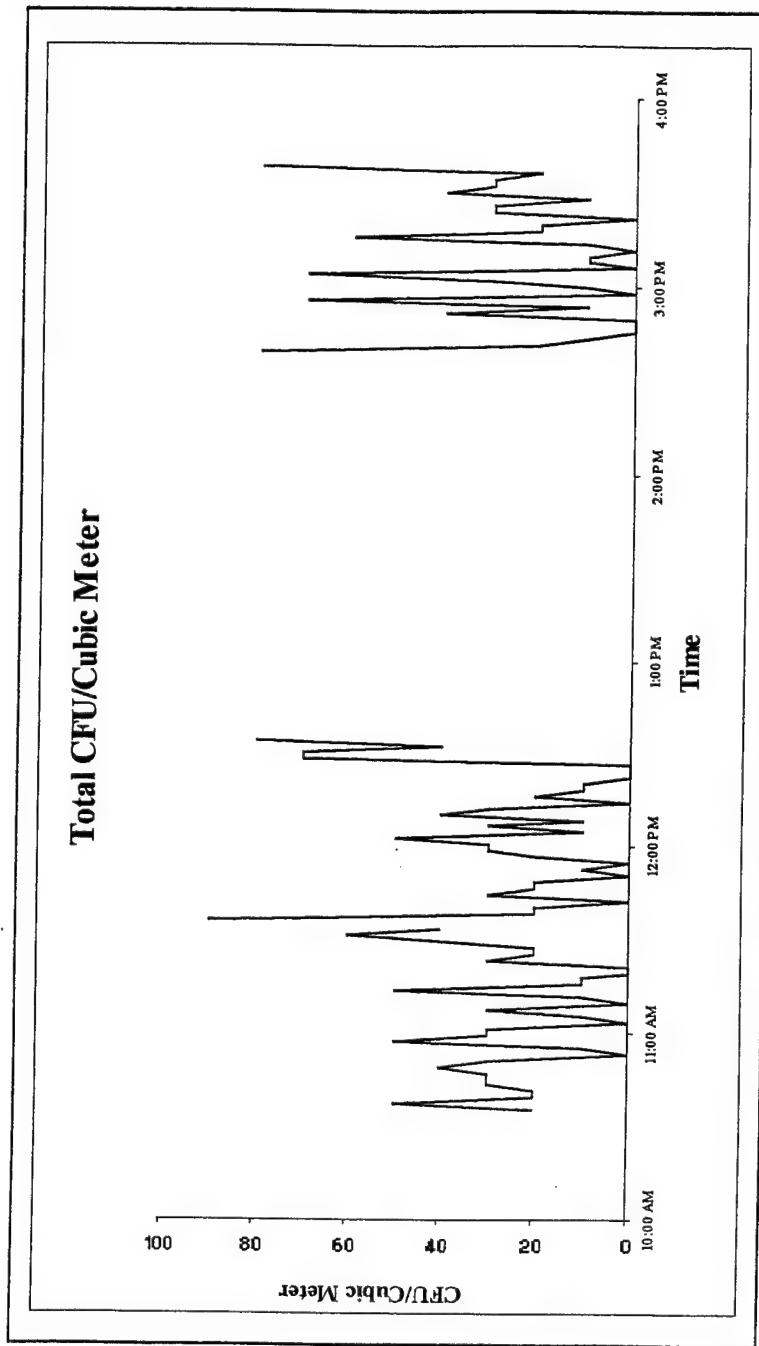


Figure 6. Example of Cultureable Bacteria Concentration in the Air.
(Note that only 1%-10% of the total bacteria in the respirable air can be cultured.)

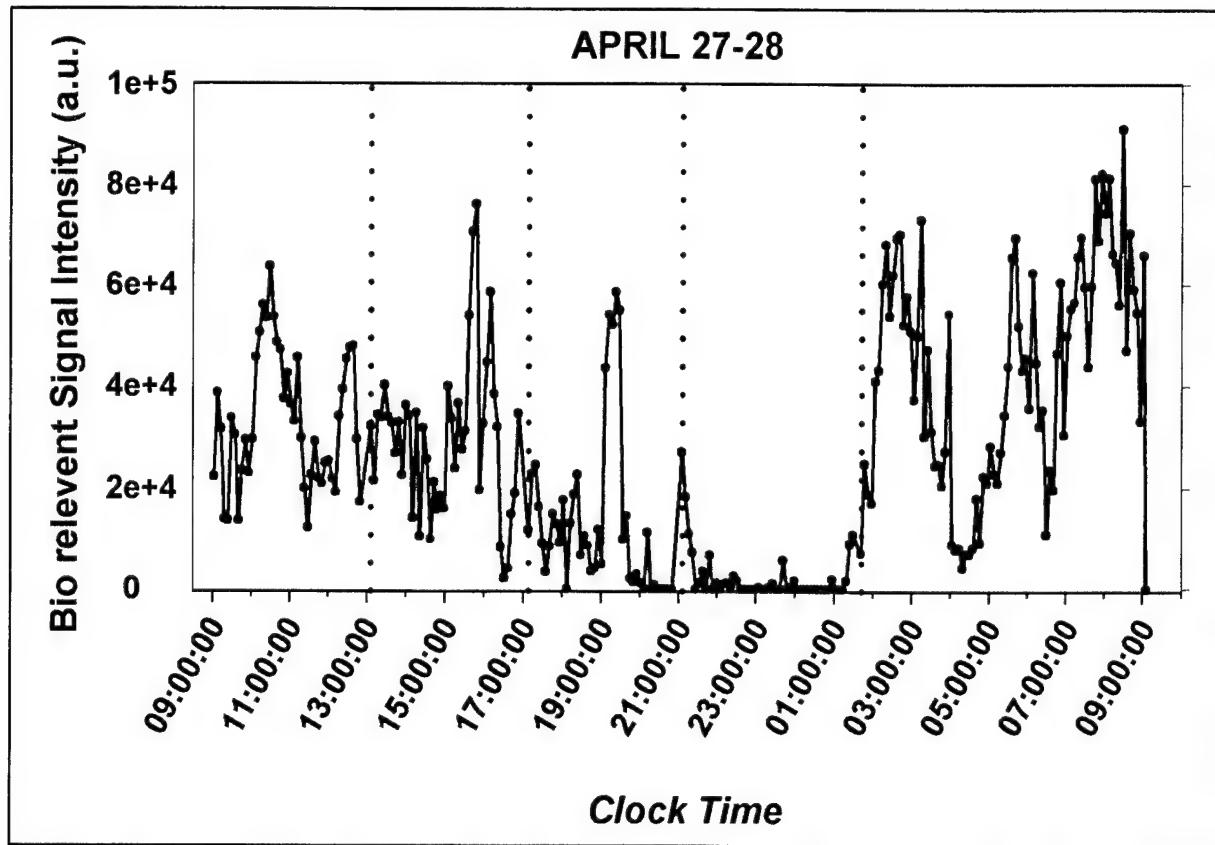


Figure 7. Example of a Py-GC/IMS (Developmental BW Detector at ECBC) Signature in the Background
(Graph provided by courtesy of Dr. Peter Snyder, ECBC)

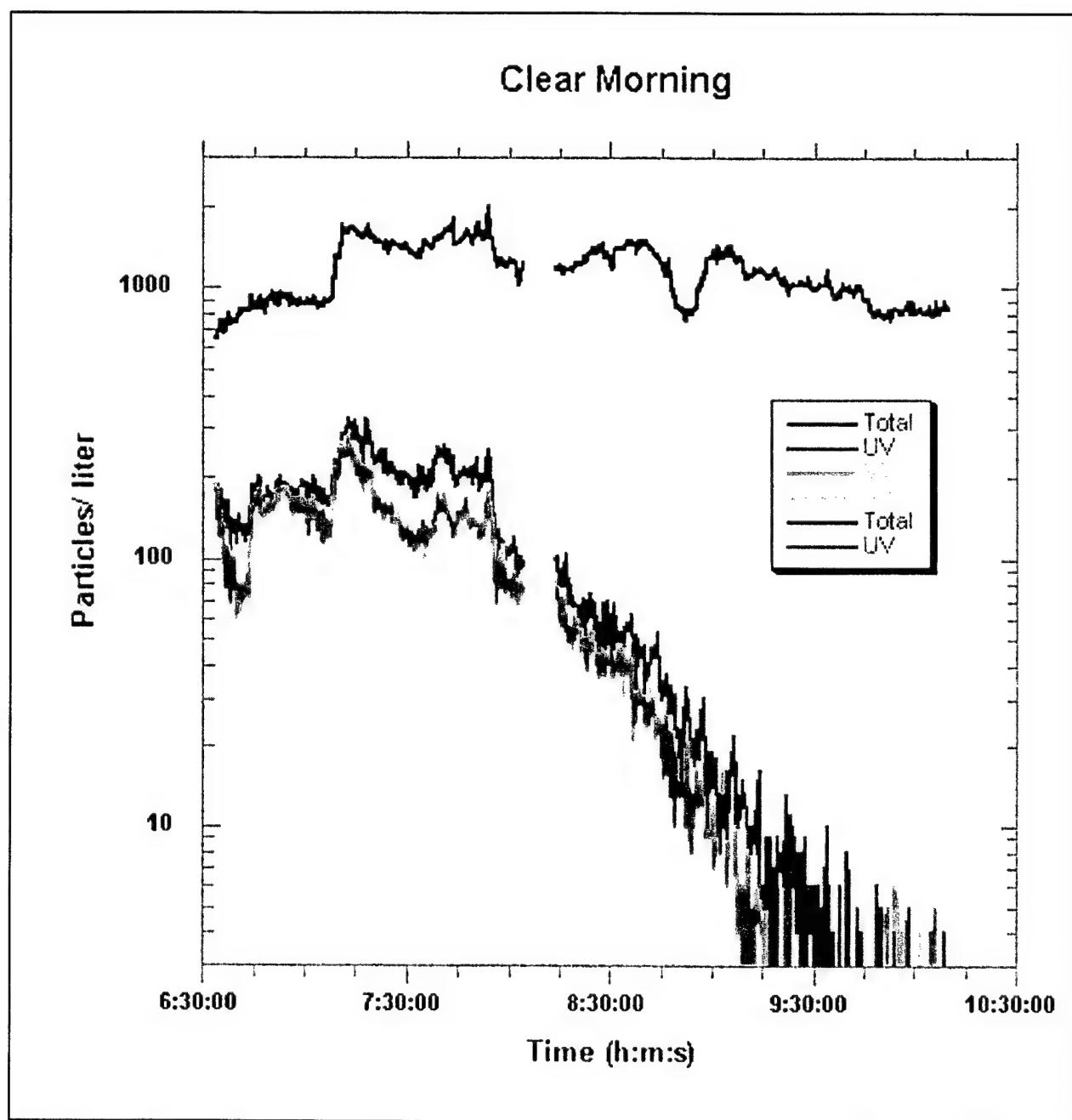


Figure 8. Example of the SPFA Signature in the Background
(Graph provided by courtesy of D. Jay Everso, NRL).

4. CONCLUSIONS AND RECOMMENDATIONS

Because of their high virulence, the detection requirements for BW agents are challenging. In general, most people cite requirements of 10 Agent Containing Particles per Liter of Air (ACPLA). As obvious from Figures 4, 6, and 8, ten ACPLA is within the natural variability of background bioaerosols.

Biological Warfare (BW) detectors are developed and tested for their ability to detect BW agents in the clean laboratory. Field tests are conducted during the Joint Field Trials (JFT). Usually the JFT are conducted at Dugway Proving Ground (DPG), DPG, UT, which is a high desert pristine area with a very low indigenous bioaerosols' background.

Normally, when BW agent detectors operate in a less pristine area rich in bioaerosols, they generate a large number of false alarms. False alarms are the nimbus of the system, and too many render such a system impotent. The main factor for reducing false alarms is the high cost in unit readiness when the alarm is given. Typically, troops at risk assume a higher Mission Oriented Protective Posture (MOPP) status, a situation that reduces their ability to fight. How long troops can remain in MOPP is further influenced by the temperature and humidity. Field commanders take a dim view of a high false alarm rate because of these considerations and the impact on unit mission. Hence, it is very important to limit the false alarm rate to a minimum.* Frequently, the material developer increases the "threshold" value for sounding the alarm. This has the effect of increasing the probability that a low-level attack will go undetected. In other words, it increases the rate of false negatives, which to a field commander is even worse.

The primary investigator used a number of respirable aerosol counters (2 – 10 μm diameter) to monitor the spatial distribution of these aerosols. He was able to show that when the counters were placed far enough apart from each other (300 - 500 m) such that their data were not correlated, and using the ensemble of counters as a unified system, he could detect a low level simulated attack by fusing the individual counters data^{15,16,17} with very low rates of false positives and false negatives. Common sense and statistical theory indicate that similar results could be obtained by using several collocated detectors that use orthogonal technologies. The principal question that needs to be resolved is: which combination of technologies could provide the best solution?

To answer this question, the authors of this report recommend that BW sensors be evaluated in the real world environment. The authors also recommend that a "background test center" be established in a location that has an active bioaerosol background (e.g., a suburban area that is surrounded or in the vicinity of fields and woods). Sensors should be brought to this background test center for evaluation early in the development stage (even at the "bread board" stage). The evaluation should be performed for extended periods (preferably for a period of at least 1 year). The rate of false positives could be established for the particular technology singly and in combination with other technologies. To evaluate the probability of detection, the

*Since 9-11, there has been increased concern of bioterrorist attacks. Limiting the false alarms to absolute minimum is even more important in the civilian arena than it is during military operations.

investigators recommend using modeling and simulation. Detector response to a given threat can be simulated and superimposed onto a real background using various discriminating algorithms.

We believe this approach will enable management to down select the best suite of detectors. Furthermore, it will provide a tool to evaluate cost benefit ratios for evaluating new technologies early in the development cycle.

Blank

LITERATURE CITED

1. Lighthart, B.; and Mohr, A.J.; Atmospheric Microbial Aerosols, Theory and Applications, Chapman and Hill, New York, NY, 1994.
2. Cox, C.S.; The Aerobiological Pathway of Microorganisms, John Wiley and Sons, New York, NY, 1987.
3. Cox, C.S.; and Wathes, C.M. (Eds.); Bioaerosols Handbook, Lewis Publishers, London, England, 1995.
4. Preece, T.P.; and Dickinson, C.H.; Ecology of Leaf-Surface Microorganisms, Academic Press, New York, NY, 1971.
5. Bernstein, M.E.; Howard, H.M.; and Caroll, G.C.; "Fluorescence Microscopy of Douglass Fir Foliage Epiflora," an. J. Microbiology, Volume 19, p 1129 (1973).
6. Fulton, J.D.; and Mitchell, R.B.; "Microorganisms of the Upper Atmosphere; II. Microorganisms in Two Types of Air Masses at 689 Meters Over a City," Appl. Microbiol. Volume 13, p 232 (1966).
7. Meuzelaar, H.L.C.; Dworzanski, J.P.; Mangoo, K.; Tripathi, A.; Cole, P.; Arnold, N.S.; Snyder, A.P.; and Maswadeh, W.; "Advances in CB Detection by Fully Automated Pyrolysis-Gas Chromatography/Ion Mobility Spectrometry (PY-GC/IMS)" (AD- E491 622). In Proceedings of the 1997 ERDEC Scientific Conference on Chemical and Biological Defense Research, ERDEC-SP-063 (AD-A356 165), July 1998.
8. Snyder, A.P.; Masqadeh, W.M.; Parsons, J.A.; Tripathi, A.; Meuzelaar, H.L.C.; Dworzanski, J.P.; and Mangoo, K.; "Field Detection of *Bacillus* Spore Aerosols with Stand-alone Pyrolysis-Gas Chromatography-Ion Mobility Spectrometry," Field Anal. Chem. and Tech. Volume 3, pp 315-326 (1999).
9. Eversole, J.D.; Hardgrove, J.; Cary, W.K.; Choulas, D.P.; and Seaver, M.; "Continuous, Rapid Biological Aerosol Detection with the Use of UV Fluorescence: Outdoor Test Results," Field Anal. Chem. and Tech. Volume 3, pp 249-259 (1999).
10. Seaver, M.; Eversole, J.D.; Hardgrove, J.J.; Cary, W.K.; and Roselle, D.C.; "Size and Fluorescence Measurements for Field Detection of Biological Aerosols," J. Aerosol Sci. and Tech. Volume 30, pp 174-185 (1999).
11. Eversole, J.D.; Cary, W.K.; Scotto, C.S.; Pierson, R. S.; and Campillo, A. J.; "Continuous Bioaerosol Monitoring Using UV Excitation Fluorescence: Outdoor Test Results," Field Analytical Chem. and Tech. Volume 15, pp 205-212 (2001).

12. Seaver, M.; Roselle, D.C.; Pinto, J.F.; and Eversole, J.D.; "Absolute Emission Spectra From *Bacillus sutilis* and *Escherichia coli* Vegetative Cells in Solution," Appl. Opt. Volume 37, pp 5344-5347 (1998).

13. Faris, G.W.; Copeland, R.A.; Mortelmans, K.; and Bronk, B.V.; "Spectrally Resolved Absolute Fluorescence Cross Sections for *Bacillus* Spores," Appl. Opt. Volume 36, pp 958-967 (1997).

14. Nachman, P.; Chen, B.; Pinnick, R.G.; Hill, S.C.; Chang, R.K.; Mayo, M.W.; and Fernandez, G.L.; "Conditional-Sampling Spectrograph Detection System for Fluorescence Measurements of Individual Biological Particles," Appl. Opt. Volume 35, pp 1069-1075 (1996).

15. Birenzvige, A.; ERDEC-TR-462, U.S. Army Edgewood Research, Development and Engineering Center, Aberdeen Proving Ground, MD, April 1998, CLASSIFIED Report.

16. Birenzvige, A.; ERDEC-TR-463, U.S. Army Edgewood Research, Development and Engineering Center, Aberdeen Proving Ground, MD, April 1998, CLASSIFIED Report.

17. Birenzvige, A.; ERDEC-TR-532, U.S. Army Edgewood Research, Development and Engineering Center, Aberdeen Proving Ground, MD, November 1998, CLASSIFIED Report.